

Extraction and Identification of Phytochemicals in N-hexane and Methanol Extracts of *Cocos Nucifera L.* Leaves

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Abstract The *Cocos nucifera L.* (Coconut palm) of Arecaceae family is found mainly in coastal areas. The extraction and analysis of various parts of plants reveal its application in natural medicine. In the present investigation, the preliminary phyto-chemical screening of *Cocos nucifera L.* leaves of Veraval region was performed using n-hexane and methanol solvents, which showed the presence of secondary metabolites like, phenols, alkaloids, tannins, terpenoids, phenols, etc. GC-MS (Gas Chromatography–Mass Spectrometry) analysis was done and Mass spectra of these compounds were matched with NIST (National Institute of Standards and Technology) library and were identified by comparing the peak area and retention time with literature and the interpretation was done. This shows presence of 11 bioactive compounds in n-hexane extract including mainly Squalene, Hexadecanoic acid, Moretenol, alpha-Tocopherol, Hexadecamethyl heptasiloxane, Eicosamethyl cyclodecasiloxane and others while the methanolic extract showed 9 bioconstituents like Squalene, Neophytadiene, Trans Phytol, A'-Neogammacer-22(29)-en-3-one and others. These bioconstituents show various biological activities like Antitumor, Antioxidant, Hypocholesterolemic, Immunostimulant, Antimycobacterial, Antiprotozoal, Anti-inflammatory, Antidepressant, Anti-inflammatory, Hypocholesterolemic, Nematicide, Pesticide, Lubricant, Neuroprotective activities and others. Thus further studies and isolation of these compounds may benefit in

pharmaceutical and medicinal fields.

Keywords Cocos Nucifera L., GC-MS, Phytochemicals, Bioactive Compounds

1. Introduction

Natural materials wide applications have been found since long for its therapeutic purposes in India. Ancient ayurvedic preparations included the use of various portions of medicinal plants, which is still relevant today and increasing popularity as people become more health-conscious. Natural medicines are preferred by millions of people in the current phase due to their effectiveness, low toxicity, and overall health benefits [1]. Natural products are also biodegradable, making them environmentally beneficial. Plants have bioactive elements in their roots, leaves, fruits, inflorescence, stems, flowers, and other parts of their bodies that have a variety of physiological functions [2], [3]. Alkaloids, tannins, saponins, terpenoids, phytols, quinones, steroids, glycosides, fatty acids, and other phytochemicals found in plants have become increasingly important in the medical sector [4]–[7]. Plant extracts are widely used as antiviral, antiallergic, antifungal, antimicrobial, antibacterial, antioxidant, anti-inflammatory, antidiabetic,

antihypertensive, and others [8].

Cocos nucifera L. (The coconut palm) that belongs to the Arecaceae family, is planted in coastal locations. After the Philippines and Indonesia, India is the world's third-largest producer. Coconut is called as the "tree of life" since all of its parts have uses ranging from industrial to household to therapeutic. According to Indian mythology, it is known as a kalpvruksh or kalpataru (wish-granting tree) and offers livelihood to people in a variety of ways. The phytochemicals found in hexane and methanolic extracts of *Cocos nucifera L.* leaves, as well as their medicinal applications, are discussed in this study. Various chemical components present in plant extracts were identified using qualitative analysis tests and GC-MS (gas chromatography combined with mass spectrometry) analysis.

2. Materials & Method

Sample collection

Coconut leaves were collected from Veraval, Gir Somnath district, Saurashtra, Gujarat during the month of December, 2021. The coconut variety selected for the investigation is Dwarf variety. The leaves were cleaned thoroughly with water and allowed to shade dry for a week and then it was powdered using grinder.

Extraction of leaves

10g of the powdered leaves were allowed to soak in

solvents like methanol and n-hexane for 24 hours. The extracts were then filtered using Whatman No 41 filter paper. Before filtering, the filter paper was moistened with methanol and hexane. Further, the GC-MS analysis and qualitative analysis for the phytochemicals screening were performed for both extracts.

Phytochemical screening

Presence and identification of various secondary metabolites like phenols, alkaloids, quinones, saponins, tannins, steroids, terpenoids, flavonoids, amino acid were done using various tests as per the standard methods in literature. Qualitative analysis was performed in both n-hexane and methanol extracts for the identification as shown in Table 1.

GC-MS analysis

Gas Chromatography-Mass Spectrometry analysis was carried out on a ShimadzuQP2010 GC-MS instrument employed with BPX capillary column at 70 eV. Carrier gas helium at a constant flow of 1.4 ml/min was maintained. 1µl of the extract was injected at a temperature maintained to 250°C and an ion source. The oven temperature was programmed to 250°C. Total GC running time was 26 min. Peaks obtained in Mass Spectrometry were compared with Wiley and NIST (National Institute of Standard and Technology) compound library.

Table 1. Qualitative Phytochemical screening in n-hexane and methanol extracts of *Cocos nucifera L.*

Preliminary Phytochemical Screening		
Tests for phytoconstituents	Results in n-hexane	Results in Methanol
Alkaloids (Mayer test)	+	+
Alkaloids (Wagner test)	+	+
Saponins	-	+
Tannins	+	+
Steroids	-	+
Terpenoids	+	+
Flavonoids	-	+
Phenols	+	+
Quinone	-	+
Amino acid	-	-

Note: + present, - absent

3. Results

Secondary metabolites in plants are responsible for various bioactivities of plants. The first step is preliminary phytochemical screening (as shown in Table 1) to determine the presence of such chemical compounds. In the present investigation, qualitative analysis of leaves of *Cocos nucifera L.* shows the positive results for alkaloids analyzed by Wagner and Mayer tests for both the hexane and methanol extracts. Formation of green colour confirmed the presence of tannins in both the extracts. Furthermore, appearance of brown colour in both the extracts shows the presence of terpenoids. However, flavonoids, saponins, steroids and quinones were found to be absent in hexane extract. Green coloured solution with

ferric chloride solution shows presence of phenols in both extracts. Tests for amino acids were found to be negative for both extracts.

GC-MS analysis of n-hexane extract of Dwarf variety showed presence of 11 phytoconstituents whereas 9 phytoconstituents were found in methanolic extract. Table 2 and Table 3 show the details of the components identified respectively for the extracts of hexane and methanol by comparing the mass spectra and retention time with the Wiley and NIST library data [9]–[11] showing its structure, molecular formula, molecular weight, name, area percentage and retention time. Figure 1 and Figure 2 show the GC-MS chromatogram of phytochemicals found in extracts of methanol and n-hexane leaves of *Cocos nucifera L.*

Table 2. Phytoconstituents identified by GC-MS analysis of n-hexane leaf extract of *Cocos nucifera L.*

Sr. No.	R. T	Area%	Name of Compound	Molecular formula	Mol. weight
1.	13.544	0.39	Hexadecamethyl heptasiloxane	C ₁₆ H ₄₈ O ₆ Si ₇	532
2.	14.155	1.97	Hexadecanoic acid	C ₁₆ H ₃₂ O ₂	256
3.	14.598	0.53	Eicosamethyl cyclodecasiloxane	C ₂₀ H ₆₀ O ₁₀ Si ₁₀	740
4.	15.161	1.58	2-Hexadecen-1-ol, 3,7,11,15-tetramethyl-, [R-[R*,R*-(E)]]- (CAS)	C ₂₀ H ₄₀ O	296
5.	15.264	0.61	Cis-Octadec-9-enal	C ₁₈ H ₃₄ O	266
6.	15.566	0.39	Tetracosamethyl cyclododecasiloxane	C ₂₄ H ₇₂ O ₁₂ Si ₁₂	888
7.	16.023	10.68	alpha-Tocopherol	C ₂₉ H ₅₀ O ₂	430
8.	17.529	0.69	Hexadecamethyl heptasiloxane	C ₁₆ H ₄₈ O ₆ Si ₇	532
9.	20.732	1.24	1H-Purin-6-amine, [(2-fluorophenyl)methyl]	C ₁₂ H ₁₀ F N ₅	243
10.	21.468	55.84	Squalene	C ₃₀ H ₅₀	410
11.	24.115	23.74	Moretenol	C ₃₀ H ₅₀ O	426

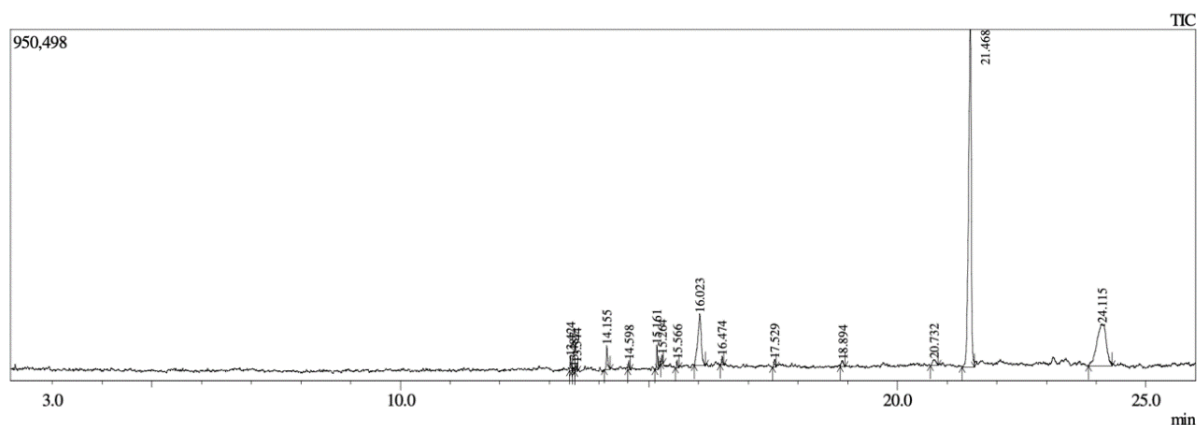
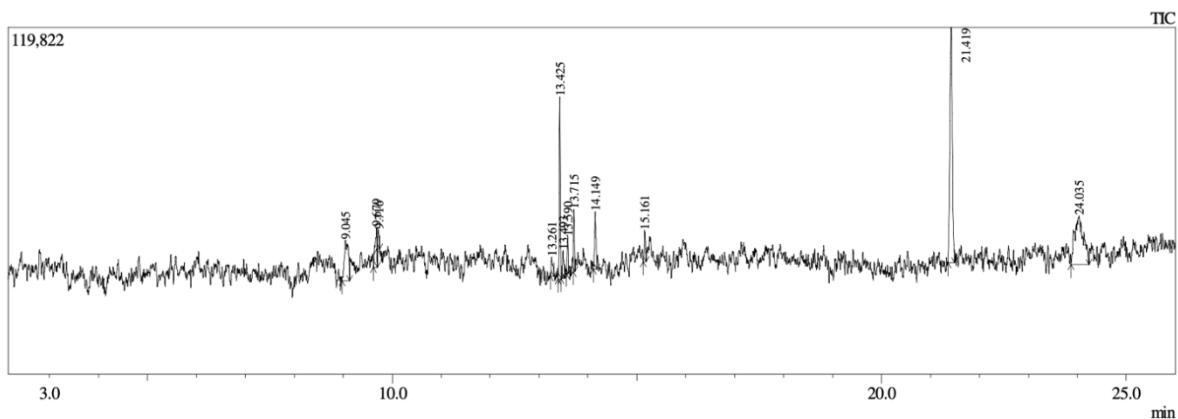


Figure 1. GC-MS Chromatogram of Phytochemicals in n-hexane leaf extract of *Cocos nucifera L.*

Table 3. Phytochemicals identified by GC-MS analysis of methanol leaf extract of *Cocos nucifera L.*

Sr. No.	R. T	Area%	Name of Compound	Molecular formula	Mol. weight
1.	9.045	9.06	Ethanone, 1-(2-hydroxy-5-methylphenyl)	C ₉ H ₁₀ O ₂	150
2.	13.425	22.73	Neophytadiene	C ₂₀ H ₃₈	278
3.	13.495	3.49	2-Hexadecene, 3,7,11,15-tetramethyl-, [R-[R*,R*-(E)]]	C ₂₀ H ₄₀	280
4.	13.590	4.59	2-Hexadecen-1-ol, 3,7,11,15-tetramethyl-, [R-[R*,R*-(E)]]	C ₂₀ H ₄₀ O	296
5.	13.715	7.75	Neophytadiene	C ₂₀ H ₃₈	278
6.	14.150	7.20	Hexadecanoic acid	C ₁₆ H ₃₂ O ₂	256
7.	15.160	3.87	2-Hexadecen-1-ol, 3,7,11,15-tetramethyl-, [R-[R*,R*-(E)]]	C ₂₀ H ₄₀ O	296
8.	21.420	29.45	Squalene	C ₃₀ H ₅₀	410
9.	24.035	6.08	A'-Neogammacer-22(29)-en-3-one (Hopeneone b)	C ₃₀ H ₄₈ O	424

**Figure 2.** GC-MS Chromatogram of Phytochemicals in methanol leaf extract of *Cocos nucifera L.*

4. Discussion

As shown in Table 4 and Table 5, the triterpenoids like Squalene identified in n-hexane (55.84% peak area, R.T. 21.468) as well methanolic (29.45% peak area, R.T. 21.420) extracts are found to have Antitumor, Antibacterial, Antioxidant, Pesticide, Cancer preventive, Lipoxigenase inhibitor, Hypocholesteremic, Immunostimulant activities. Hexadecanoic acid (1.97% peak area, R.T. 14.155) in n-hexane and (7.20% peak area, R.T. 14.150) in methanol extract, is a saturated fatty acid that exhibits Antioxidant, Anti-cancer, Hypocholesterolemic, 5-alpha reductase, Nematicide, Lubricant, inhibitors activity, Pesticide, Antiandrogenic, Flavor, Hemolytic, 5-Alpha reductase inhibitor activities. Vitamin E (alpha tocopherol) (16.023% peak area, R.T. 10.68) in n-hexane extract is an immunity booster and acts as an Antioxidant. Moretenol (23.74% peak area, R.T. 24.155), a triterpenoid found in n-hexane extract possesses Antimycobacterial, Antiprotozoal, Anti-inflammatory activities. Neophytadiene, a sesquiterpenoids is found to show Antimicrobial,

Antivenom, Antidepressant, Anti-inflammatory and Neuroprotective activities in methanol extract with 22.73% peak area at 13.425 R.T. Ethanone, 1-(2-hydroxy-5-methylphenyl) belonging to Alkyl Phenyl Ketone class acts as 17-beta-hydroxysteroid dehydrogenase-Inhibitor, Testosterone-Hydroxylase-Inducer and Aryl-Hydrocarbon-Hydroxylase-Inhibitor having peak area of 9.06% and 9.045 R.T. A'-Neogammacer-22(29)-en-3-one (Hopeneone b) in methanolic extract have been reported as Hypocholesterolemic and antioxidant at 6.08 R.T and 24.0359 % peak area. Trans Phytol (2-Hexadecen-1-ol, 3,7,11,15-tetramethyl-, [R-[R*,R*-(E)]] identified in both n-hexane (1.58% peak area, R.T. 15.161) as well as methanol (3.87% peak area, R.T. 15.160) extract is a Neuroprotective, Antimicrobial, Anti-inflammatory, Anticancer, Anti-diabetic, Antioxidant, Anti-diuretic properties, Antidepressant, Anticonvulsant. Therefore, n-hexane and methanol extracts of leaves of *Cocos nucifera L.* are found to possess various biological activities and recommended for further studies.

Table 4. Reported Biological activity and class of compounds for phytochemicals identified in hexane leaf extract of *Cocos nucifera* L.

Sr. No.	Name of the Compounds	Class of Compounds	Reported Biological activity	Reference
1.	Hexadecamethyl heptasiloxane	Organosiloxane	Antibacterial, Antimicrobial	[12], [13]
2.	Hexadecanoic acid	Palmitic acid (Saturated fatty acid)	Antioxidant, Antiandrogenic, Nematicide, Hypocholesterolemic, Pesticide, Lubricant, Flavor, Hemolytic, 5-Alpha reductase inhibitor activities	[5], [14], [15]
3.	Eicosamethyl cyclodecasiloxane	Organohetero silanes	Antimicrobial, Antihelminthic, Antioxidant	[16]
4.	Trans Phytol [2-Hexadecen-1-ol, 3,7,11,15-tetramethyl-, [R-[R*,R*-(E)]]-]	Acyclic diterpenoids	Antimicrobial, Anti-inflammatory, Anticancer, Anti-diabetic, Antioxidant, Anti-diuretic properties, Antidepressant, Anticonvulsant, Neuroprotective	[17]–[22]
5.	Cis-Octadec-9-enal	Fatty aldehyde	Flavouring agent	[23], [24]
6.	Tetracosamethyl cyclododecasiloxane	Organohetero silanes	Hepatoprotective, Antispasmodic, Antirheumatic	[16]
7.	alpha-Tocopherol	Vitamins	Immunity Booster, Antioxidant	[25]–[27]
8.	Hexadecamethyl heptasiloxane	Organosiloxanes	Antimicrobial	[16]
9.	1H-Purin-6-amine, [(2-fluorophenyl)methyl]	Alkaloids or Purines	Antimicrobial, as a potent mechanism-based inhibitor of several enzymes like cholesterol acyltransferase, acyl coenzyme A, monoamine oxidase, Antifungal	[28], [29]
10.	Squalene	Triterpenoids	Antitumor, Antibacterial, Antioxidant, Pesticide, Cancer preventive, Lipoygenaseinhibitor, Hypocholesteremic, Immunostimulant	[30], [31]
11.	Moretenol	Triterpenoids	Antimycobacterial, Antiprotozoal, Anti-inflammatory	[32], [33]

Table 5. Reported Biological activity and class of compounds for phytochemicals identified in methanol leaf extract of *Cocos nucifera* L.

Sr. No.	Name of the Compounds	Class of Compounds	Reported Biological activity	Reference
1.	Ethanone, 1-(2-hydroxy-5-methylphenyl)	Alkyl Phenyl Ketone	17-beta-hydroxysteroid dehydrogenase-Inhibitor, Testosterone-Hydroxylase-Inducer, Aryl-Hydrocarbon-Hydroxylase-Inhibitor,	[16]
2.	Neophytadiene	Sesquiterpenoids	Antimicrobial, Antivenom. Antidepressur Anti-inflammatory and Neuroprotective activities.	[34], [35]
3.	2-Hexadecen-1-ol, 3,7,11,15-tetramethyl-, [R-[R*,R*-(E)]]	Acyclic diterpenoids	Antimicrobial, Anti-inflammatory, Anticancer, Anti-diabetic, Antioxidant, Anti-diuretic properties, Antidepressant, Anticonvulsant, Neuroprotective	[17]–[22]
4.	Hexadecanoic acid	Palmitic acid (Saturated fatty acid)	Antioxidant, Lubricant, Anti-cancer, Hypocholesterolemic, 5-alpha reductase, Pesticide, inhibitors activity, Antiandrogenic, Nematicide, pFlavor, Hemolytic, 5-Alpha reductase inhibitor activities.	[36]
5.	Squalene	Triterpenoids	Antitumor, Antibacterial, Antioxidant, Pesticide, Cancer preventive, Lipoygenaseinhibitor, Hypocholesteremic, Immunostimulant	[30], [31]
6.	A'-Neogammacer-22(29)-en-3-one (Hopeneone b)	Hopanoids	Antioxidant, Hypocholesterolemic	[37], [38]

5. Conclusion

The preliminary phytochemical screening gives the basic information regarding the presence of compounds in the plants. In the present study, the hexane and methanolic extracts of *Cocos nucifera L.* revealed the positive tests for alkaloids, tannins, terpenoids, and phenols shown in Table 1. Further analysis using GC-MS of hexane and methanol extract of leaves of *Cocos nucifera L.* showed the presence of significant bioactive constituents like Squalene, Hexadecanoic acid, Moretenol, alpha-Tocopherol, Hexadecamethyl heptasiloxane, Eicosamethyl cyclodecasiloxane, Neophytadiene, Trans Phytol, 2-hexadecen-1-ol, 3,7,11,15-tetramethyl-[R-[R*,R*-(E)]], A'-Neogammacer-22(29)-en-3-one and others. These constituents exhibited important biological activities like Antitumor, Antioxidant, Immunostimulant, Antimycobacterial, Antiprotozoal, Anti-inflammatory, Antidepressant, Nematicide, Hypocholesterolemic, Pesticide, Neuroprotective, Lubricant activities and others. They should be further explored for various pharmaceutical and pharmacological applications.

REFERENCES

- [1] R. Yadav and M. Agarwala, "Phytochemical analysis of some medicinal plants," *Journal of Phytology*, vol. 3, no. 12, pp. 10–14, 2011.
- [2] B. E. Ogeyemhe, R. A. Amaechi, C. D. Ekpruke, B. O. Airiagbonbu, and E. B. Odigie, "Comparative benefits of *Cocos nucifera L.* husk, milk and shell extracts on body weight changes and haematological indices in male rats," *Tropical Journal of Natural Product Research*, vol. 4, no. 8, pp. 455–462, 2020, doi: 10.26538/tjnpr/v4i8.22.
- [3] V. Palengara, Dhanya G, and Ashisg GR, "Phytochemical analysis of coconut shell (*Cocos nucifera* Linn.) using gas chromatography-mass spectrometry (GC-MS)," *J Pharmacogn Phytochem*, vol. 7, no. 6, pp. 384–386, 2018.
- [4] M. Parimalam, V. Pushpa Rani, and K. Deepak Kumar, "Study of Phytochemical Screening and GC-MS Analysis of two Extracts of Coconut *Haustorium*," *J Adv Sci Res*, vol. 12, no. 2, pp. 348–353, 2021.
- [5] Elaiyaraja A and Chandramohan G, "Comparative phytochemical profile of *Indoneesiella echioides* (L.) Nees leaves using GC-MS A Elaiyaraja and G Chandramohan," *J Pharmacogn Phytochem*, vol. 5, no. 6, pp. 158–171, 2016.
- [6] R. Kavitha, "Phytochemical Screening and GC-MS Analysis of Bioactive Compounds present in Ethanolic Extracts of leaf and fruit of *Trichosanthesis Dioica* Roxb.," *Article in International Journal of Pharmaceutical Sciences and Research*, vol. 12, no. 5, p. 2755, 2021, doi: 10.13040/IJPSR.0975-8232.12(5).2755-64.
- [7] V. Hooda, G. N. Sharma, N. Tyagi, and A. Hooda, "Phytochemical and Pharmacological Profile of *Cocos nucifera* : An Overview," *International Journal of Pharmacy & Therapeutics*, vol. 3, no. 2, p. 130, 2012.
- [8] R. Karadi, P. Parekh, A. Shah, and P. Azmi, "Antimicrobial Activities of *Musa paradisiaca* and *Cocos nucifera* & evaluation of gelling potentials of *Hibiscus cannabinus* seed mucilage," *Article in International Journal of Research in Pharmaceutical and Biomedical Sciences*, 2011.
- [9] Y. Massada, "Analysis of essential oil by gas chromatography and spectrometry," *John Wiley & Sons*, New York, NY, 334., 1976.
- [10] L. G. Johnsen, P. B. Skou, B. Khakimov, and R. Bro, "Gas chromatography – mass spectrometry data processing made easy," *J Chromatogr A*, vol. 1503, pp. 57–64, Jun. 2017, doi: 10.1016/J.CHROMA.2017.04.052.
- [11] L. M. Egerton-Warburton and E. L. Ghisalberti, "Essential oil composition of *Chamelaucium uncinatum*," *Phytochemistry*, vol. 40, no. 3, pp. 837–839, Oct. 1995, doi: 10.1016/0031-9422(95)00328-5.
- [12] S. M. Mohy El-Din and A. M. D. El-Ahwany, "Bioactivity and phytochemical constituents of marine red seaweeds (*Jania rubens*, *Corallina mediterranea* and *Pterocladia capillacea*) ScienceDirect Bioactivity and phytochemical constituents of marine red seaweeds (*Jania rubens*, *Corallina mediterranea* and *Pterocladia capillacea*)-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>)," *Journal of Taibah University for Science*, vol. 10, pp. 471–484, 2016, doi: 10.1016/j.tjusc.2015.06.004.
- [13] A. M. Syeda and K. Riazunnisa, "Data on GC-MS analysis, in vitro anti-oxidant and anti-microbial activity of the *Catharanthus roseus* and *Moringa oleifera* leaf extracts," *Data Brief*, vol. 29, Apr. 2020, doi: 10.1016/J.DIB.2020.105258.
- [14] Ponnamma S.U. and Manjunath K., "GC-MS analysis of phytochemicals in the methanolic extract of *Justicia wynaadensis* (Nees) T. Anders.," *International Journal of Pharma and Biosciences*, 2012.
- [15] H. Malipeddi and M. Das, "Phytochemical screening, GC-MS analysis and biological activity of *Ipomoea eriocarpa* leaf extracts," *Article in International Journal of Pharmacy and Pharmaceutical Sciences*, vol. 6, no. 4, pp. 592–594, Mar. 2014.
- [16] Dr. Duke's Phytochemical and Ethnobotanical Databases, "Dr. Duke's Phytochemical and Ethnobotanical Databases," U.S. Department of Agriculture, Agricultural Research Service. 1992-2016. 2016.
- [17] G. Rajeswari, M. Murugan, and V. R. Mohan, "GC-MS Analysis of Bioactive Components of *Hugonia Mystax L.* (Linaceae)," *Research Journal of Pharmaceutical and Bio Chemical Science*, vol. 3, no. 4, pp. 301–308, 2012.
- [18] N. Hamidi, L. Ziane, M. Djellouli, and HA. Lazouni, "Chemical characterization by gc-ms from the aerial parts of *fagonia longispina* (Zygophyllaceae)," *Asian Journal of Pharmaceutical and Clinical Research*, vol. 9, no. 1, pp. 175–176, 2016.
- [19] H. Phukan, C. R. Bora, and P. K. Mitra, "Phytochemical Screening and GC-MS Analysis of Methanolic leaf Extract of an Endemic Plant *Kayea assamica*," *IOSR Journal of Pharmacy and Biological Sciences (IOSR-JPBS)*, vol. 12, no. 5, pp. 7–16, 2017, doi: 10.9790/3008-1205020716.
- [20] A. N. Syad, B. S. Rajamohamed, K. P. Shunmugaiah, and P. D. Kasi, "Neuroprotective effect of the marine macroalga

- Gelidiella acerosa: identification of active compounds through bioactivity-guided fractionation,” *Pharm Biol*, vol. 54, no. 10, pp. 2073–2081, Oct. 2016, doi: 10.3109/13880209.2016.1145700.
- [21] J. Costa et al., “Evaluation of Antioxidant Activity of Phytol Using Non- and Pre-Clinical Models,” *Curr Pharm Biotechnol*, vol. 17, no. 14, pp. 1278–1284, Nov. 2016, doi: 10.2174/1389201017666161019155715.
- [22] W. Lee, E. R. Woo, and D. G. Lee, “Phytol has antibacterial property by inducing oxidative stress response in *Pseudomonas aeruginosa*,” *Free Radic Res*, vol. 50, no. 12, pp. 1309–1318, Dec. 2016, doi: 10.1080/10715762.2016.1241395.
- [23] Henry Rakoff, “Synthesis of unsaturated aldehydes,” *J Am Oil Chem Soc*, vol. 46, no. 6, pp. 277–279, 1969, doi: <https://doi.org/10.1007/BF02545003>.
- [24] E. Makowicz, I. Jasicka-Misiak, T. Dariusz, and P. Kafarski, “Botanical Origin Authentication of Polish Phacelia Honey Using the Combination of Volatile Fraction Profiling by HS-SPME and Lipophilic Fraction Profiling by HPTLC,” *Chromatographia*, vol. 82, pp. 1541–1553, 2019, doi: 10.1007/s10337-019-03778-x.
- [25] W. A. Skinner and R. M. Parkhurst, “Antioxidant Properties of α -Tocopherol Derivatives and Relationship of Antioxidant Activity to Biological Activity,” *Lipids*, vol. 5, no. 2, pp. 184–186, 1970.
- [26] J. F. Mallet, C. Cerrati, E. Ucciani, J. Gamisans, and M. Gruber, “Antioxidant activity of plant leaves in relation to their α -tocopherol content,” *Food Chem*, vol. 49, no. 1, pp. 61–65, Jan. 1994, doi: 10.1016/0308-8146(94)90233-X.
- [27] G. Marques, A. Gutiérrez, and J. C. del Río, “Chemical characterization of lignin and lipophilic fractions from leaf fibers of curaua (*Ananas erectifolius*),” *J Agric Food Chem*, vol. 55, no. 4, pp. 1327–1336, Feb. 2007, doi: 10.1021/jf062677x.
- [28] V. A. Kumar, K. Ammani, B. Siddhardha, U. Sreedhar, and G. A. Kumar, “Differential biological activities of the solvent extracts of *Cerriops decandra* (Griff.) and their phytochemical investigations,” *J Pharm Res*, vol. 7, no. 7, pp. 654–660, Jul. 2013, doi: 10.1016/J.JOPR.2013.05.024.
- [29] T. P. Muthu, “Analysis of Bioactive Constituents from the Flesh of *Turbo brunneus* (Roding, 1798) By GCMS,” ~ 257 ~ *International Journal of Fisheries and Aquatic Studies*, vol. 3, no. 1, pp. 257–259, 2015.
- [30] A. Wei and T. Shibamoto, “Antioxidant activities and volatile constituents of various essential oils,” *J Agric Food Chem*, vol. 55, no. 5, pp. 1737–1742, Mar. 2007, doi: 10.1021/jf062959x.
- [31] V. Thangapandian and M. Sermakkani, “GC-MS analysis of *Cassia Italica* leaf methanol extract,” *Asian Journal of Pharmaceutical and Clinical Research*, vol. 5, no. 2, pp. 90–94, 2012.
- [32] M. Z. Pérez-González, G. A. Gutiérrez-Rebolledo, L. Yáñez-Mulía, I. S. Rojas-Tomé, J. Luna-Herrera, and M. A. Jiménez-Arellanes, “Antiprotozoal, antimycobacterial, and anti-inflammatory evaluation of *Cnidioscolus chayamansa* (Mc Vaugh) extract and the isolated compounds,” *Biomedicine & Pharmacotherapy*, vol. 89, pp. 89–97, May 2017, doi: 10.1016/J.BIOPHA.2017.02.021.
- [33] W. A. Abdel-Naime, J. R. Fahim, M. A. Fouad, and M. S. Kamel, “Antibacterial, antifungal, and GC-MS studies of *Melissa officinalis*,” *South African Journal of Botany*, vol. 124, pp. 228–234, Aug. 2019, doi: 10.1016/J.SAJB.2019.05.011.
- [34] Y. Inoue, T. Hada, A. Shiraishi, K. Hirose, H. Hamashima, and S. Kobayashi, “Biphasic effects of geranylgeraniol, teprenone, and phytol on the growth of *Staphylococcus aureus*,” *Antimicrob Agents Chemother*, vol. 49, no. 5, pp. 1770–1774, May 2005, doi: 10.1128/AAC.49.5.1770-1774.2005.
- [35] M. Bhardwaj, V. K. Sali, S. Mani, and H. R. Vasanthi, “Neophytadiene from *Turbinaria ornata* Suppresses LPS-Induced Inflammatory Response in RAW 264.7 Macrophages and Sprague Dawley Rats,” *Inflammation*, vol. 43, no. 3, pp. 937–950, Jun. 2020, doi: 10.1007/s10753-020-01179-z.
- [36] Yu et al., “Isolation and characterization of methyl esters and derivatives from *Euphorbia kansui* (Euphorbiaceae) and their inhibitory effects on the human SGC-7901 cells,” 2005.
- [37] S. Arora and G. Kumar, “Phytochemical screening of root, stem and leaves of *Cenchrus biflorus* Roxb,” *J Pharmacogn Phytochem*, vol. 7, no. 1, pp. 1445–1450, 2018.
- [38] M. Akbar et al., “*Cornus macrophylla*, the Antibacterial Activity of Organic Leaf Extracts and the Characterization of the More Lipophilic Components by GC/MS,” *Molecules*, vol. 25, no. 10, pp. 1–11, 2020, doi: 10.3390/molecules25102395.