

Development and Standardization of a Simple and Quick Screening Protocol for Arsenic Phyto-toxicity Tolerance at Seedling Stage in Rice

Md. Abu Syed^{1,*}, K. M. Iftokharuddaula¹, Md. Golam Rasul², G. K. M. Mustafizur Rahmam³,
Golam M. Panaullah⁴, John M. Duxbury⁵, Mohammad Hossain⁶, Partha S. Biswas¹

¹Plant Breeding Division, Bangladesh Rice Research Institute (BRRI), Bangladesh

²Department of Genetics and Plant Breeding, Bangabandhu Sheikh Mujibur Rahman Agricultural University (BSMRAU), Bangladesh

³Department of Soil Science, Bangabandhu Sheikh Mujibur Rahman Agricultural University (BSMRAU), Bangladesh

⁴Former Director Research, Bangladesh Rice Research Institute (BRRI), Bangladesh

⁵Department of Crop and Soil Sciences, Cornell University, USA

⁶Plant Pathology Division, Bangladesh Rice Research Institute (BRRI), Bangladesh

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Abstract Cultivation with Arsenic (As) contaminated water severely affects growth of rice plants that results into reduced grain yield. This study was carried out to develop a rapid and effective method of screening rice varieties and/or segregating population against arsenic phyto-toxicity tolerance. Sodium arsenate was used as arsenic source. Ten pre-germinated seeds of each variety were grown on styro-foam placed on a tray filled with either phosphate-free nutrient solution (control) or the same nutrient solution supplemented with di-sodium hydrogen arsenate (treatment). The trays were kept at controlled environmental condition in a growth chamber at $26\pm 1^\circ\text{C}$ with 70% humidity and 3500 lux. After 7 days of growth, standard growth parameters such as shoot length, root length, root-shoot biomass were recorded and percentages reduction for shoot length, root length and root shoot biomass compared to those of control were calculated. Arsenic stress reduced the phenotypic expression of all three seedling traits but the effect of arsenic was more prominent on root length compared to the shoot length and root-shoot biomass. The dose response experiment showed that the 0.75-1.25 mgL^{-1} Arsenic concentrations had much promise to discriminate rice genotypes for arsenic phyto-toxicity tolerance at early growth stage. The validation experiment with the 0.75-1.25 mgL^{-1} Arsenic doses on a set of 20 genotypes identified 1.25 mgL^{-1} Arsenic could effectively and efficiently differentiate rice genotypes, particularly for tolerance in shoot length and root-shoot biomass reduction, where 1.0 mgL^{-1} Arsenic was found sufficient for discriminating varieties for tolerance in root length reduction. The rice variety BRRI dhan47 showed more

tolerance against Arsenic contaminated water.

Keywords Arsenic, Hydroponic, Reactive Oxygen Species, Least significant Difference, Arsenic Phyto-toxicity, Rice

1. Introduction

Arsenic (As), a non-essential element for plant growth, it is generally phyto-toxic and is expected to negatively affect plant growth [1-3]. As a result of many negative effects, arsenic causes reduced growth of plants. The primary response of plants is the generation of reactive oxygen species (ROS) after exposure to arsenic. ROS can directly damage proteins, amino acids and nucleic acids and cause peroxidation of membrane lipids [4]. Arsenic accumulates in the plant tissue and stimulates peroxidase synthesis during the early phases of plant development, long before the appearance of visible changes [5-7]. Rice yield and biomass production are greatly reduced at elevated Arsenic concentrations, often leading to death of plant population [8-10]. Long-term irrigated soils subjected to Arsenic contaminated ground water may create a significant risk to animal and human health via soil-crop transfer of Arsenic. In addition, consumption of excess Arsenic from groundwater water has caused human health problems in several countries [11-14]. Therefore, many researchers in the entire world have shown keen interest in arsenic toxicity, screening and selecting arsenic tolerant and remediative plants and mechanisms related to

their detoxification.

Since there are good evidences that metal tolerance is under genetic control [17]. To facilitate breeding programmes, a rapid, non-destructive, inexpensive and repeatable seedling-based bioassay is required for selection of tolerant genotypes from early segregating generations [18]. Field screening methodology is less precise, less efficient and, therefore, less attractive to breeders since this method is challenged by many factors, including soil heterogeneity in physical and chemical characteristics, tedious excavations of roots, low screening rate, and time-consuming when a large number of genotypes are under evaluation. Therefore, a rapid and effective screening system is needed to discriminate tolerant and sensitive genotypes. Hydroponics, on the other hand, is more attractive than the soil-based screening because it allows rapid evaluation of a large number of genotypes in a shorter time on the basis of relative growth rate and toxicity [19]. Besides, accurate phenotypic data are expected from hydroponics because of the tight control of the screening conditions. Importantly, hydroponics enables observations to be made of intra- and inter-specific genetic variation in plant responses, in terms of levels of tolerance shown and the specific tolerance mechanisms that are employed. The use of hydroponic culture has been suggested as a means of assessing the plant tolerance to the toxic elements or the efficiency in mineral utilization [20-22]. Hydroponic techniques have been utilized by many scientists to identify and understand the tolerance mechanism under excess heavy metals in plants. Meharg [23] reported hydroponic culture as a suitable way to screen and identify heavy metal tolerant and remedial plants before field studies.

Improving arsenic tolerance in rice plants is the most suitable way to decrease destructive effects of arsenic toxicity [24-25]. Since tolerance is genetically controlled, screening and selection of Arsenic tolerant rice genotypes at seedling stage under hydroponics condition can lead to the development of tolerant cultivars/varieties against excess arsenic because hydroponics provides easy access to root system, exact control over available nutrients and pH and non-destructive measurements of tolerance. But, rapid screening procedure for arsenic phyto-toxicity

tolerance in rice is available only for root length parameter [20]. But only root length parameter is not sufficient for arsenic tolerant variety development. Three growth parameters (root length, shoot length and root-shoot biomass) were considered in this study. Considering the above fact the present study has been undertaken with the following objectives: i) to develop and standardize a quick and short-cut screening protocol; ii) to discriminate rice varieties and/or segregating population into tolerant and susceptible groups effectively and efficiently with a single dose of arsenic treatment under controlled condition.

2. Materials and Methods

The experiment was conducted at the screening house of Plant breeding field Laboratory, Bangladesh Rice Research Institute (BRRI), Gazipur-1701.

2.1. Plant Materials

Two arsenic tolerant (BRRI dhan47 and BRRI dhan54) and two sensitive rice varieties (BRRI dhan29 and BRRI dhan45) identified from previous study [26] were used in dose response experiment. Another twenty arsenic tolerant and sensitive rice varieties including some IRRI lines, US and Chinese cultivars were used for validation of identified in the dose response experiment. The seeds were collected from the Plant Breeding Division and Gene Bank of Bangladesh Rice Research Institute (BRRI), Gazipur-1701.

2.2. Experimental Design and Treatments

Dose response experiment The experiment was laid out in a Randomized Complete Block Design (RCBD). The treatments were arranged in a 4 x 7 factorial scheme comprising four rice genotypes (BRRI dhan29, BRRI dhan45, BRRI dhan47 and BRRI dhan54) and seven concentrations of arsenic in solution (0, 0.25, 0.50, 0.75, 1.0, 1.25 and 1.50 mg L⁻¹ Arsenic) with four replications. Layout of dose response experiment was as below:

V4 As5 V4 As4 V2 As1 V4 As3 V2 As3 V3 As6 V3 As3 V3 As2 V4 As7 V4 As2 V4 As6 V1 As6 V2 As7 V1 As5 V3 As5 V4 As1 V1 As7 V1 As4 V2 As2 V3 As4 V1 As1 V1 As3 V3 As1 V2 As6 V2 As4 V1 As2 V2 As5 V3 As7	Rep1
V4 As2 V1 As6 V3 As5 V3 As3 V2 As5 V2 As6 V3 As7 V1 As4 V1 As5 V2 As3 V3 As4 V1 As3 V1 As2 V1 As7 V4 As5 V4 As4 V4 As7 V2 As4 V2 As7 V3 As6 V2 As1 V4 As3 V3 As2 V4 As1 V3 As1 V2 As2 V4 As6 V1 As1	Rep2
V1 As1 V2 As3 V4 As2 V3 As5 V3 As7 V4 As7 V3 As2 V3 As3 V2 As5 V3 As4 V1 As4 V1 As3 V4 As6 V1 As2 V2 As1 V2 As2 V1 As6 V2 As6 V1 As7 V3 As6 V2 As4 V3 As1 V1 As5 V4 As3 V2 As7 V4 As4 V4 As5 V4 As1	Rep3
V2 As1 V1 As7 V3 As3 V4 As2 V4 As6 V3 As2 V3 As4 V2 As6 V1 As5 V1 As2 V1 As3 V4 As7 V3 As6 V1 As6 V2 As3 V2 As5 V2 As2 V3 As1 V2 As7 V4 As3 V4 As4 V2 As4 V3 As5 V3 As7 V1 As4 V4 As1 V4 As5 V1 As1	Rep4

Legend: V1=BRRI dhan29, V2=BRRI dhan45, V3= BRRI dhan47 and V4=BRRI dhan54;
As1= 0, As2=0.25, As3=0.50, As4=0.75, As5=1.0, As6=1.25 and As7=1.50 mg L⁻¹ Arsenic

Dose validation experiment The experiment for dose validation with 20 genotypes was conducted for 3 levels of Arsenic (0.75, 1.0 and 1.25 mg L⁻¹) in three replications. Arsenic (As) was used in the form of NaH₂AsO₄ (Analytical reagent grade; BDH). The phosphate-free nutrient solution consisted of 0.1 mM Mg²⁺ and SO₄²⁻, 0.2 mM Ca²⁺ and K⁺ and 0.6 mM NO₃⁻ [27] was used to grow seedlings. Layout of dose validation experiment was given below:

V1 As2 V5 As1 V12 As1 V20 As1 V7 As3 V9 As1 V6 As3 V1 As3 V12 As2 V2 As2 V15 As2 V10 As1 V13 As3 V3 As1 V8 As3	Rep1
V16 As3 V17 As3 V18 As1 V4 As1 V18 As2 V2 As1 V18 As3 V13 As1 V6 As1 V2 As3 V14 As1 V10 As3 V6 As2 V8 As2 V9 As2	
V19 As3 V4 As2 V3 As2 V10 As2 V7 As2 V19 As1 V19 As2 V13 As2 V15 As1 V9 As3 V4 As3 V14 As3 V20 As2 V5 As3 V17 As2	
V15 As3 V14 As2 V16 As2 V8 As1 V1 As1 V20 As3 V11 As1 V5 As2 V12 As3 V11 As3 V7 As1 V11 As2 V3 As3 V17 As1 V16 As1	Rep2
V15 As1 V13 As3 V7 As1 V6 As1 V4 As3 V8 As3 V3 As3 V17 As3 V6 As3 V3 As1 V8 As1 V9 As1 V5 As1 V2 As3 V10 As2	
V2 As2 V9 As3 V11 As2 V6 As2 V16 As2 V9 As2 V14 As1 V11 As3 V12 As2 V1 As3 V17 As1 V16 As1 V8 As2 V3 As2 V18 As3	
V20 As3 V11 As1 V18 As2 V15 As2 V1 As1 V4 As2 V1 As2 V7 As2 V15 As3 V5 As3 V2 As1 V13 As1 V13 As2 V20 As1 V4 As1	Rep3
V20 As2 V10 As3 V7 As3 V17 As2 V19 As2 V14 As2 V19 As3 V5 As2 V12 As1 V12 As3 V14 As3 V16 As3 V18 As1 V19 As1 V10 As1	
V14 As2 V20 As1 V1 As3 V1 As1 V3 As3 V19 As3 V16 As1 V12 As1 V17 As2 V4 As3 V6 As3 V7 As2 V7 As1 V19 As2 V15 As1	
V20 As2 V1 As2 V9 As2 V16 As3 V10 As1 V18 As2 V14 As3 V3 As1 V10 As3 V8 As2 V15 As3 V20 As3 V3 As2 V13 As3 V2 As2	Rep3
V5 As3 V9 As1 V18 As3 V12 As3 V8 As1 V2 As1 V4 As1 V6 As2 V18 As1 V14 As1 V7 As3 V4 As2 V6 As1 V8 As3 V11 As2	
V17 As3 V16 As2 V11 As1 V5 As1 V5 As2 V17 As1 V15 As2 V13 As1 V10 As2 V12 As2 V2 As3 V9 As3 V11 As3 V19 As1 V13 As2	

Legend: V1=IR44595, V2=IR9209-26-2, V3=Tie 90-1, V4=Aijiaonante 100, V5=Cocodire, V6=Priscilla, V7=Jing 185-7, V8=Jinnvo No. 6, V9=BRR1 dhan28, V10=BRR1 dhan29, V11=BRR1 dhan45, V12=BRR1 dhan47, V13=HUA565, V14=HUA564, V15=Weed tolerant Rice, V16=BRR1 dhan50, V17=BRR1 dhan61, V18=BRR1 dhan49, V19=Jefferson and V20=BRR1 dhan54; As1=0.75, As2=1.0 and As3=1.25 mg L⁻¹ Arsenic

2.3. Experimental Set-Up

Seeds were disinfected with Vitavex for 2 hours and soaked overnight in petridishes after washing with distilled water and then incubated at 30°C for 48 hours. Ten pre-germinated seeds were placed on styrofoam floats with a mosquito net placed at the bottom. Then the floats with pre-germinated seeds were placed on the tray filled with either phosphate-free nutrient solution (control) or the same nutrient solution supplemented with di-sodium hydrogen arsenate (treatment). The trays were kept in a growth chamber at 26 ± 1°C with 70% humidity and 3500 lux, white fluorescent light (55 μmol m⁻²s⁻¹). After 7 days of growth, shoot length (cm) was measured from the base of the culm to the tip of the longest leaf and root length (cm) was recorded from the root-shoot junction to the tip of the longest root. Root and shoot samples were dried in an oven at 50°C for 96 hours and then weighed to determine their biomass (mg).

2.4. Statistical Analysis

The rate of reduction in shoot length, root length and root-shoot biomass was estimated as the percentage of reduction in shoot length, root length and root shoot biomass, respectively compared to control following a formula given as,

$$\text{Rate of reduction (\%)} = 1 - (\text{Growth with arsenic} - \text{Growth without arsenic}) * 100$$

The data were subjected to analyses of variance (ANOVA) to determine the significance of individual effects (genotypes and arsenic concentration) and genotype x Arsenic treatment interactions. Least significant differences (LSD) were calculated at P = 0.05 for comparisons between mean values. The ANOVA and LSD test were performed using Statistix 10 software (Tallahassee, FL, USA) and rate of reduction, standard deviation (SD), range, coefficient of variation (CV) were calculated using Excel 2007.

3. Results

The analysis of variance (ANOVA) of arsenic dose response and dose validation experiments revealed significant variation in shoot length reduction, root length reduction and root-shoot reduction among various genotypes and different arsenic concentrations. The interaction effects were also significant (Table 1).

Table 1. Mean squares of the analysis of variance for shoot length reduction (%), root length reduction (%), root-shoot biomass reduction (%) in dose response experiment

Sources	Degree of freedom	Reduction (%)		
		Shoot length	Root length	Root-shoot biomass
Genotypes (G)	3	887.51***	3949.27***	250.98***
Concentrations (C)	5	3740.06***	1997.00***	2470.68***
G x C	15	17.57***	35.57***	36.02NS
Error	69	5.15	5.02	21.61
Coefficient of variation (%)		27.24	21.45	36.12
Heritability (h ² b)		97.72	99.49	72.63

*** Significant at 0.1% level of significance, ^{NS} Non-significant

3.1. Shoot Length Reduction

Genotypic mean comparison showed that the lowest shoot length reduction (47.59%) was in BRRI dhan47 which was statistically different from other genotypes. The maximum shoot length reduction was observed in the genotype BRRI dhan54 (61.74%), followed by BRRI dhan29 (58.19%), and BRRI dhan45 (57.66%) (Table 2). Concentration means revealed that shoot length reduction was increased under increasing of Arsenic concentrations and was also significantly differed from each other. It was maximum (71.69%) at 1.50 mgL⁻¹ followed by 1.25 mgL⁻¹

(64.54%). Shoot length reduction was very close and statistically similar at 1.00 mgL⁻¹ (62.18%) to that at 0.75 mgL⁻¹ (61.43%) level of applied arsenic. The lower reduction was found at 0.25 mgL⁻¹ (28.91%) followed by 0.50 mgL⁻¹ (49.01%). The shoot length reduction among genotypes except BRRI dhan47 was almost close to each other at 0.25 mg L⁻¹, 0.50 mg L⁻¹ and 1.50 mg L⁻¹ levels of arsenic. But at the concentration from 0.75 mg L⁻¹ to 1.25 mg L⁻¹ of Arsenic, the genotypic difference was apparent and significant. The range along with standard deviation was also the highest at the Arsenic concentration of 0.75 mg L⁻¹ to 1.25 mg L⁻¹.

Table 2. Mean of shoot length reduction (%) over different concentration (mg L⁻¹) of arsenic in dose response experiment

Genotypes	Arsenic concentration (mg L ⁻¹)						Genotypic means
	0.25	0.50	0.75	1.0	1.25	1.50	
BRRI dhan29	27.9 bc	49.6 a	64.7ab	65.3 b	66.7 b	74.8 a	58.2
BRRI dhan45	29.4 ab	52.2 a	61.8 b	62.4 b	66.6 b	73.6 a	57.7
BRRI dhan47	24.5c	41.5 b	50.3 c	51.6 c	54.8 c	62.9 b	47.6
BRRI dhan54	33.7 a	52.8 a	68.9a	69.5 a	70.0 a	75.5a	61.8
Concentration means	28.91	49.01	61.43	62.18	64.54	71.69	
LSD (0.05)	1.60						1.31
Standard Deviation	3.82	5.22	7.97	7.66	6.69	5.92	
Range	24.5-33.7	41.5-52.8	50.3-68.9	51.6-69.5	54.8-70.0	62.9-75.4	

Within column different letters indicate significance differences among rice varieties (LSD test: P< 0.05)

Table 3. Mean shoot length reduction (%) over three levels of arsenic concentrations (mg L⁻¹) of 20 rice genotypes

Genotypes	Arsenic concentration (mg L ⁻¹)			Genotypic Mean
	0.75	1.0	1.25	
IR44595	46.06 ij	51.55 kl	56.94 ghi	51.51
IR9209-26-2	59.54 b	65.69 ab	68.26 b	64.51
Tie 90-1	56.07 bc	60.29 d	64.20 c	60.18
Aijiaonante 100	49.15 ghi	57.20 efg	59.76 ef	55.39
Cocodire	64.61 a	55.97 fgh	61.45 de	57.33
Priscilla	53.24 cdefg	54.43 hij	56.77 ghi	54.81
Jing 185-7	55.71 bc	57.34 ef	61.98 d	58.34
Jinvo No. 6	44.89 j	48.58 m	52.44 j	48.67
BRRI dhan28	50.17 fghi	52.70 jk	59.94 ef	54.26
BRRI dhan29	59.67 b	63.98 bc	67.70 b	63.68
BRRI dhan45	52.05 cdefg	55.89 fgh	58.41 fg	55.46
BRRI dhan47	39.03 k	44.87 n	46.86 k	43.58
HUA565	54.87 cd	55.33 ghi	56.14 hi	55.43
HUA564	51.52 defgh	53.59 ij	58.41 fg	54.53
Weed tolerant Rice	51.54 defgh	56.47fg	57.57 gh	55.22
BRRI dhan50	47.52 hij	50.22 lm	55.51 i	51.07
BRRI dhan61	53.71 cdef	66.73 a	73.93 a	64.76
BRRI dhan49	54.06 cdef	-	-	54.07
Jefferson	50.49 efg	59.12 de	62.09 d	57.20
BRRI dhan54	-	56.04 fgh	62.73 cd	59.41
Concentration means	52.41	56.42	60.27	
LSD value at 0.05	0.66			1.63
Standard deviation	5.68	5.59	5.93	
Range	39.03-64.61	44.87-66.73	46.86-73.93	

Within column different letters indicate significance differences among rice varieties (LSD test: P< 0.05)

Validation trial with 0.75 mg L⁻¹, 1.0 mg L⁻¹ and 1.25 mg L⁻¹ Arsenic over a set of 20 diverse genotypes showed that the minimum mean shoot length reduction was with BRRI dhan47 (43.58%) followed by Jinnvo no. 6 (48.67%), while the maximum mean reduction was found with BRRI dhan61 (64.76%) followed by IR9209-26-2 (64.51%) and BRRI dhan29 (63.68%) (Table 3). The shoot length reduction was increased with increasing arsenic concentration. The highest mean reduction (60.27%), the highest standard deviation (5.93) and the largest range (46.86-73.93) in shoot length reduction were observed at 1.25 mg L⁻¹ Arsenic among all the Arsenic concentrations. The highest difference in shoot length reduction between the known tolerant variety (BRRI dhan47) and sensitive varieties (BRRI dhan29 and BRRI dhan54) was also found at 1.25 mg L⁻¹ level of Arsenic compared to other concentrations.

3.2. Root Length Reduction

Genotypic mean of root length reduction was found least in BRRI dhan47 (53.57%), which was statistically different from other genotypes in dose response experiment. Rice varieties BRRI dhan45 (80.35%) and BRRI dhan29 (79.44%) showed the highest root length reduction that was statistically similar followed by BRRI dhan54 (77.75%)

(Table 4). Concentration means showed that root length reduction was the minimum at 0.25 mg L⁻¹ and the maximum at 1.50 mg L⁻¹ although all concentration means were statistically different from each other. The maximum statistical differentiation among genotypes was found at 1.25 mg L⁻¹ followed by 1.50 mg L⁻¹, 1.0 mg L⁻¹ and 0.25 mg L⁻¹. The maximum differentiation at 0.25 mg L⁻¹ and 1.50 mg L⁻¹ doses were actually varietal difference probably due to lower and higher dose. The highest range and standard deviation were observed at the Arsenic concentration of 0.75 mg L⁻¹ followed by 0.5 mg L⁻¹ and 1.25 mg L⁻¹.

In validation trial, BRRI dhan47 exhibited significantly least root length reduction (58.89%) among 20 genotypes (Table 5). Among three levels of arsenic, the highest mean root length reduction (78.75%) was found at 1.25 mg L⁻¹ Arsenic. The differences in percent of root length reduction between BRRI dhan47 and BRRI dhan45, and BRRI dhan47 and BRRI dhan54 were the maximum at 1.00 mg L⁻¹ Arsenic but between BRRI dhan47 and BRRI dhan29, it was at 0.75 mg L⁻¹. The highest standard deviation (7.83) between the genotypes was found at 0.75 mg L⁻¹ Arsenic, although 1.00 mg L⁻¹ Arsenic had the reduction rate ranging from (59.77%-87.56%), which was almost similar to that of 1.25 mg L⁻¹ Arsenic.

Table 4. Mean of root length reduction (%) over different concentration (mg L⁻¹) of arsenic in dose response experiment

Genotypes	Arsenic concentration (mg L ⁻¹)						Genotypic means
	0.25	0.50	0.75	1.0	1.25	1.50	
BRRI dhan29	54.4 b	78.8 a	80.0 a	85.3 a	87.1 b	91.0 a	79.4
BRRI dhan45	61.7 a	75.8 a	79.6 a	84.8 a	89.6 a	90.6 ab	80.3
BRRI dhan47	37.8 c	47.3 b	48.7 b	57.1 c	58.7 d	71.8c	53.6
BRRI dhan54	59.9 a	74.3 a	77.6 a	81.4 b	83.7 c	89.5 b	77.8
Concentration means	53.47	69.06	71.50	77.15	79.79	85.71	
LSD (0.05)	1.58						1.29
Standard Deviation	10.92	14.63	15.22	13.46	14.27	9.31	
Range	37.8-61.7	47.3-78.8	48.7-80.0	57.1-85.3	58.7-89.6	71.8-91.0	

Within column different letters indicate significance differences among rice varieties (LSD test: P< 0.05)

Table 5. Mean root length reduction (%) over three level of arsenic concentrations (mg L^{-1}) of 20 rice genotypes

Genotypes	Arsenic concentration (mg L^{-1})			Genotypic means
	0.75	1.0	1.25	
IR44595	68.92 ef	80.76 bcd	84.30 b	77.99
IR9209-26-2	73.15 cdf	81.59 b	83.49 b	79.81
Tie 90-1	64.13 gh	70.87 i	72.41 hi	69.06
Aijiaonante 100	56.01 i	70.99 i	74.35 g	67.14
Cocodire	74.59 bcd	80.98 bc	84.21 b	79.86
Priscilla	71.59 def	79.39 cdef	81.00 cde	77.34
Jing 185-7	76.31 bc	78.97 def	81.49 cd	78.97
Jinnvo No. 6	72.45 cde	76.23 g	79.70 de	76.13
BRR1 dhan28	56.61 i	68.21 j	70.81 ij	65.22
BRR1 dhan29	75.96 bc	77.43 fg	79.78 de	77.72
BRR1 dhan45	67.93 fg	79.53 cde	80.63 de	76.00
BRR1 dhan47	55.80 i	59.77 l	61.35 k	58.89
HUA565	75.65 bcd	78.06 efg	79.51 e	77.77
HUA564	63.18 h	64.50 k	69.07 j	65.62
Weed tolerant Rice	76.72 bcd	80.28 bcd	81.22 cde	79.42
BRR1 dhan50	82.85 a	87.56 a	88.86 a	86.45
BRR1 dhan61	74.55 bcd	87.05 a	88.90 a	83.44
BRR1 dhan49	78.04 b	-	-	78.04
Jefferson	75.70 bcd	81.15 bc	82.72 bc	79.89
BRR1 dhan54	-	74.13 h	77.37 f	75.71
Concentration means	70.82	76.38	78.75	
LSD at 0.05		0.68		1.65
Standard deviation	7.83	7.11	6.81	
Range	55.80-82.85	59.77-87.56	61.35-88.90	

Within column different letters indicate significance differences among rice varieties (LSD test: $P < 0.05$)

3.3. Root-Shoot Biomass Reduction

Genotypic mean of root-shoot biomass reduction was observed minimum (29.62%) in BRR1 dhan47 followed by BRR1 dhan45 (33.33%), BRR1 dhan54 (35.22%) and BRR1 dhan29 (37.37%) (Table 6). Concentration means revealed that root-shoot biomass reduction was the minimum at 0.25 mg L^{-1} and the maximum at 1.50 mg L^{-1} although all concentration means are statistically dissimilar from each other except 0.75 mg L^{-1} to 1.0 mg L^{-1} and at 1.0 mg L^{-1} to at 1.25 mg L^{-1} . Arsenic concentration level from 0.75 mg L^{-1} to 1.50 mg L^{-1} revealed the highest statistical differentiation among genotypes except 1.25 mg L^{-1} . The highest range of root-shoot biomass reduction among the genotypes was observed at the concentration of 0.75 mg L^{-1} followed by 1.0 mg L^{-1} , 0.5 mg L^{-1} , 1.25 mg L^{-1} and 0.25

mg L^{-1} where as, standard deviation was the highest at the concentration of 0.75 mg L^{-1} followed by 0.5 mg L^{-1} , 1.25 mg L^{-1} , 1.0 mg L^{-1} and 0.25 mg L^{-1} .

Root-shoot biomass was comparatively less affected at different doses of Arsenic in dose validation experiment. The minimum genotypic mean of root shoot-biomass reduction was observed in BRR1 dhan47 (28.79%) compared to IR9209-26-2 (47.41%) and Jing 185-7 (46.39%) (Table 7). The maximum root-shoot biomass reduction was observed at higher concentration (1.25 mg L^{-1}) of arsenic in culture solution. The difference between tolerant variety (BRR1 dhan47) and sensitive varieties (BRR1 dhan45, and BRR1 dhan29) for root-shoot biomass reduction was higher at 1.25 mg L^{-1} Arsenic. The range (31.60-52.76) and standard deviation (6.63) were also recorded high at 1.25 mg L^{-1} Arsenic.

Table 6. Mean of root-shoot biomass reduction (%) over different concentration (mg L⁻¹) of arsenic in dose response experiment

Genotypes	Arsenic concentration (mg L ⁻¹)						Genotypic means
	0.25	0.50	0.75	1.0	1.25	1.50	
BRR1 dhan29	10.0 a	30.5a	43.2 a	43.9 a	45.3 a	51.3 a	37.37
BRR1 dhan45	9.6 a	24.9a	37.5 ab	38.3 ab	40.8 a	48.8 ab	33.33
BRR1 dhan47	15.5 a	20.9 a	30.4 b	33.3 b	34.6 b	43.1 c	29.62
BRR1 dhan54	14.9 a	30.4 a	37.8 ab	38.6 ab	42.4 a	47.3 b	35.22
Concentration means	12.49	26.67	37.23	38.52	40.77	47.63	
LSD (0.05)	3.28						2.67
Standard Deviation	3.10	4.65	5.24	4.34	4.53	3.45	
Range	9.6-15.5	20.9-30.5	30.4-43.2	33.3-43.9	34.6-45.3	43.1-51.3	

Within column different letters indicate significance differences among rice varieties (LSD test: P< 0.05)

Table 7. Mean root-shoot biomass reduction (%) over three levels of arsenic concentrations (mg L⁻¹) of 20 rice genotypes

Genotypes	Arsenic concentration (mg L ⁻¹)			Genotypic Mean
	0.75	1.0	1.25	
IR44595	25.98 fg	28.65 l	32.73 mn	29.13
IR9209-26-2	43.24 a	46.28 ab	52.75 a	47.41
Tie 90-1	28.47 efg	34.33 hij	38.34 jkl	33.72
Aijiaonante 100	29.83 defg	36.79 ghi	42.66 fgh	36.42
Cocodire	42.07 a	43.60 bcde	47.04 cde	44.08
Priscilla	35.40 bcd	38.92 fg	42.35 ghi	39.23
Jing 185-7	41.52 a	47.20 a	50.39 abc	46.39
Jinnvo No. 6	34.66 cd	36.90 gh	38.97 ijk	36.38
BRR1 dhan28	31.58 def	33.33 jk	40.14 hij	35.02
BRR1 dhan29	31.34 def	45.59 abc	49.23 bcd	42.06
BRR1 dhan45	31.90 de	44.87 abcd	49.69 abc	42.15
BRR1 dhan47	24.94 g	29.86 l	31.61 n	28.79
HUA565	32.31 de	33.54 ijk	35.74 klm	33.85
HUA564	30.54 defg	33.34 jk	36.84 jkl	33.56
Weed tolerant Rice	39.04 abc	42.08 def	45.82 def	42.34
BRR1 dhan50	26.48 efg	30.46 kl	35.32 lm	30.69
BRR1 dhan61	27.51 efg	44.00 abcde	52.29 ab	41.25
BRR1 dhan49	41.11 ab	-	-	41.12
Jefferson	31.05 def	40.79ef	44.28 efg	38.73
BRR1 dhan54	-	42.59 cde	47.34 cde	44.99
Concentration means	33.20	39.02	43.14	
LSD at 0.05	1.09			2.65
Standard deviation	5.66	6.18	6.63	
Range	24.94-43.24	28.66-47.21	31.60-52.76	

Within column different letters indicate significance differences among rice varieties (LSD test: P< 0.05)

4. Discussion

Arsenic is generally phyto-toxic and negatively affects plant growth [1], particularly yield and biomass production in rice are greatly reduced at elevated Arsenic concentrations, often leading to death of plant population [8-10]. Irrigation with Arsenic contaminated ground water greatly affects early growth stages of rice plants inhibiting phosphorus uptake from the soil. However, arsenate is an analog of inorganic phosphate (Pi) and is easily transported across the plasma lemma by Pi transporter (PHT) protein [28-29]. When phosphate competes with arsenate for uptake, arsenate toxicity is lower under high phosphate condition. Phosphate free hydroponic solutions [30] therefore, provide the clearest insights into the potency of externally supplied Arsenic on whole plant growth because arsenate is out compete phosphate for

entry into the plant [31].

During early growth stage root and shoot development, particularly, root length, shoot length and root-shoot biomass reduced significantly at elevated arsenic concentration in phosphate free culture solution compared to control. Arsenic stress reduced the phenotypic expression of all the seedling traits; shoot length, root length and root-shoot biomass (Figure 1). Although, the effect of arsenic was more prominent on root length compared to the shoot length and root-shoot biomass indicating lesser severity of arsenic on shoot growth and biomass. Abedin et al. [32] observed that plant roots were the first tissue of contact to Arsenic in nutrient media, where the metalloid inhibited root extension and proliferation. Similar adverse effect of arsenic on root length was observed by Carbonell-Barrachina et al. [33].

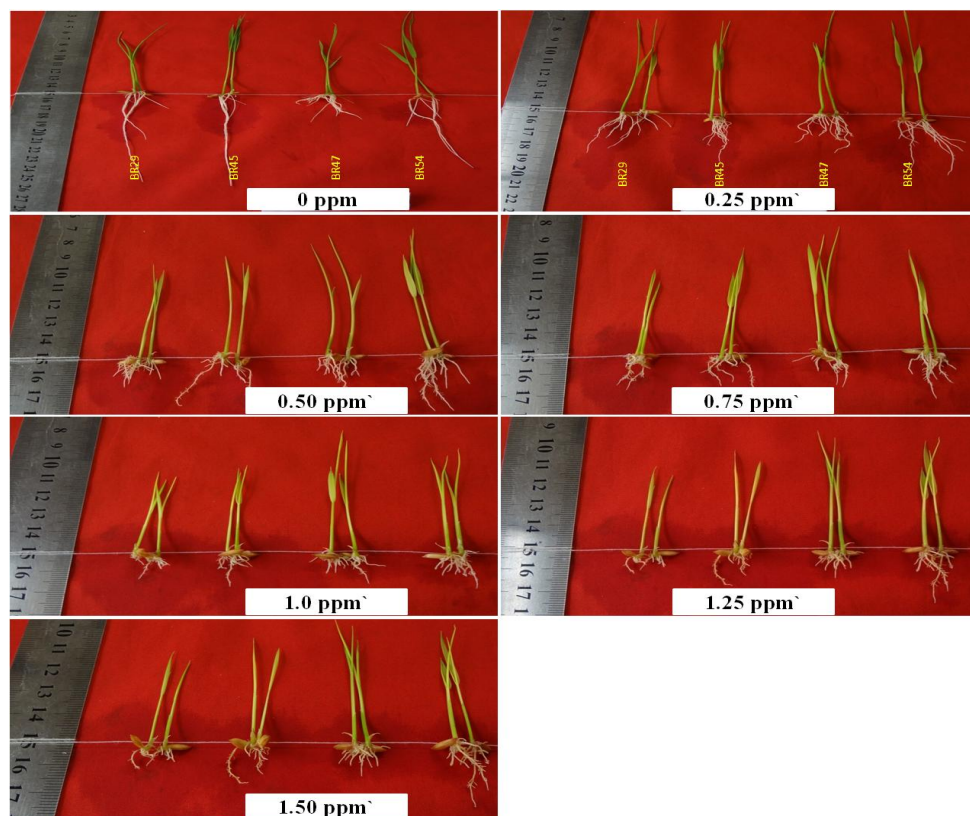


Figure 1. Images of seedling showing effect of different levels of arsenic concentration on early growth stage of different rice genotypes (BR29=BRR I dhan29, BR45=BRR I dhan45, BR47=BRR I dhan47 and BR54=BRR I dhan54, respectively, from left to right in the photographs)

The dose response experiment with six level of Arsenic starting from 0.25 mg L^{-1} to 1.5 mg L^{-1} over two tolerant varieties (BRR I dhan47 and BRR I dhan54) and two susceptible varieties (BRR I dhan29 and BRR I dhan45) showed maximum statistical differentiation, range and standard deviation at $0.75\text{-}1.25 \text{ mg L}^{-1}$ arsenic treatment for shoot length reduction, root-shoot biomass reduction but for root length reduction arsenic concentrations starting from 0.50 mg L^{-1} to 1.25 mg L^{-1} Arsenic were found highly responsive to differentiate genotypes. Considering mean reduction, standard deviation and range of the genotypes in all three growth traits, the $0.75\text{-}1.25 \text{ mg L}^{-1}$ Arsenic concentrations were found suitable to screen for arsenic tolerance in rice at early growth stage. Also the tolerant variety, BRR I dhan47 showed minimum reduction in all three growth traits at $0.75\text{-}1.25 \text{ mg L}^{-1}$ Arsenic level.

The dose validation experiment with 0.75 mg L^{-1} , 1.0 mg L^{-1} and 1.25 mg L^{-1} over a set of 20 diverse genotypes including BRR I dhan47 as tolerant check and BRR I dhan29 and BRR I dhan45 as susceptible checks also showed minimum and maximum reduction, respectively in all three growth traits. The reduction was the most acute, highly significant and maximum segregation among the genotypes was found at 1.25 mg L^{-1} Arsenic in both shoot length reduction and root-shoot biomass reduction traits. Similar trend of response for root length reduction was observed at 1.0 mg L^{-1} Arsenic concentration. Reduced root length at early growth stage under arsenic stress in the

present study was parallel to the findings of Dasgupta et al. [20] who identified $13.3 \mu\text{M}$ (i.e. 1.0 mg L^{-1}) arsenate was suitable dose to screen root length reduction trait of diverse genotypes under hydroponic arsenic stress condition. Abedin et al. [32] reported significant reduction in root biomass at $>1 \text{ mg L}^{-1}$ Arsenic(V). Marin et al. [34] found significant reduction of rice shoot length when arsenite or monomethyl arsenic acid was applied at a dose of 0.8 mg L^{-1} . Therefore, this study suggested that 1.25 mg L^{-1} of Arsenic could be effective and efficient for discriminating tolerant and sensitive rice genotypes considering all three growth traits (root length, shoot length and root-shoot biomass) against arsenic phytotoxicity at early growth stage in hydroponic culture.

5. Conclusions

Arsenic (arsenate compound) was found to be toxic to seedling growth of rice. Root length was significantly reduced with increasing arsenic concentrations in the hydroponic solution. Effective and efficient differentiation was observed at 1.25 mg L^{-1} Arsenic for shoot length and root-shoot biomass, while 1.0 mg L^{-1} Arsenic was sufficient for root length trait. Taking into account of the above discussions, it could be concluded that the concentration of 1.25 mg L^{-1} of Arsenic was the best for detection of differences between tolerant and sensitive rice genotypes

at early growth stage considering all three growth traits (root length, shoot length and root-shoot biomass) against arsenic toxicity which would be a simple and quick screening protocol. Hence, this study would provide a good scope for identifying genotypes with higher level of tolerance to Arsenic stress from mass screening of segregating or diverse sets of genotypes. This study also further proved BRR1 dhan47's promise to be used as a potential donor in the future rice breeding programmes for arsenic phyto-toxicity tolerance.

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